Table I. Chemical Shifts and Coupling Constants for the Methyl Diazonium and Methylene Diazenium Ions

	species ^a				
	CH ₂ LOS-				
	$CH_2LN_2^+$	$CH_2N_2^+L$	O_2F	CH ₃ OH ₂ +	CH_2N_2
$\delta(^1\mathrm{H})^b$	4.75	6.09	4.21	4.7, 9.4°	3.28 ^d
$\delta(^{13}C)$	43.78	73.28	63.20	62.9 ^e	23.1 ^f
$^{2}J_{HD}^{g}$	2.2		1.6		
JHC	163.3	176.0	155.5	157.5 ^e	
J_{DC}	24.7		23.5		

^a L signifies H or D. ^b Chemical shifts were measured in parts per million relative to Me₄Si = 0. ^c Reference 6. ^d Reference 7. ^e Reference 8. f Reference 9. g Coupling constants were measured in hertz.

formed on heating this sample was monodeuterated, as witnessed by triplet resonances in its ¹H and ¹³C spectra; therefore H-D exchange and, consequently, reversible protonation on carbon is insignificant in the intermediates.

The most reasonable structures compatible with these observations are 1 for the major intermediate and 2 for the minor one; their chemical shifts and coupling constants are collected in Table I along with reference data for diazomethane, methyl fluorosulfate, and the methyl oxonium ion.

It is possible that the methyl diazonium ion be not free in solution, but rather associated with, or bonded to, the fluorosulfate counterion.¹⁰ This is considered unlikely, however, as incorporation of antimony pentafluoride (up to 50% the acid concentration) to the solution did not change the ¹H or ¹³C shifts. Furthermore, the ¹⁹F spectrum of the reaction mixture¹¹ at -100 °C showed only the peaks of SO₂ClF at -99.1 ppm and HFSO₃ at -41.7 ppm (relative to CFCl₃). After the mixture was heated to -80 °C, the resonance of methyl fluorosulfate appeared at -31.4 ppm (lit.¹² -31.2 ppm).

Protonated diazirene could be considered as an alternative structure to 2.¹⁰ It can be excluded, however, as introduction of diazirene^{13,11} into the reaction medium at -120 °C did not produce any previously observed species, but only one with ¹³C shift of 33.8 ppm (δ (¹³C) of diazirene 10.9 ppm), which was unaffected by heating the mixture to -80 °C.

In accordance with the proposed structures 1 and 2, both the ¹H and ¹³C chemical shifts of **2** are considerably greater than those of 1. Surprisingly, although the proton chemical shift of 1 is very similar to those of methyl fluorosulfate and the methyl oxonium ion, indicating similar electron distributions in the three methyl groups, the ¹³C shift of **1** is almost 20 ppm upfield from the other two, though nonetheless 20 ppm downfield from the diazomethane value. As the one-bond coupling constants ${}^{1}J_{CH}$ and ${}^{1}J_{CD}$ in Table 1 indicate that the diazonium group is slightly more electron withdrawing than the fluorosulfate or oxonium groups, this must be taken as another example of the unreliability of the ¹³C shift as a measure of electron density, at least for diazo and diazonium compounds.14

The broadening (15 Hz) of the ¹³C resonance of 2 was observed not only for the singlet in the proton decoupled spectrum, but also for the triplet in the proton coupled spectrum, and so is unlikely to derive from poor resolution of fortuitously close peaks. An alternative explanation-rapid exchange of the N-H proton-is acceptable only if confined to an intimately bound complex, as otherwise the diazomethane formed would also be reprotonated on carbon, leading to the eventual demise of the N-protonated species. A probable source of broadening for the resonance of 2 but not of 1 is a greater degree of ${}^{13}C{}^{-14}N$ coupling in the former. It has previously been shown that, whereas in diazomethane this coupling can be observed at room temperature,¹⁵ less symmetrically substituted diazoalkanes have merely broadened lines for the diazo carbons. The electrical field gradient around the central nitrogen in 1 is less symmetrical than that in 2, entailing a more rapid ¹⁴N quadrupolar relaxation. This in turn removes the ${}^{13}C{}^{-14}N$ coupling and gives a narrower line for the diazonium ion resonance.

The C-protonated species 1 has been calculated to have in the gas phase a lesser free enthalpy of formation than that of the N-protonated isomer by 125 kJ mol⁻¹ (basis set 4-31G), or 209 kJ mol⁻¹ (basis set STO-3G).¹⁶ The solvational energy assigned to an +N-H bond in fluorosulfuric acid is 46 kJ mol^{-1} ;¹⁷ so from these values 1 would be predicted to be the only observable isomer in HFSO₃-SO₂ClF. Hence, the isomer ratio of 4:1 observed in our study could indicate the relative rates of irreversible protonation on the two basic centers of diazomethane.

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References and Notes

- (1) Mohrig, J. R.; Keegstra, K. J. Am. Chem. Soc. 1967, 89, 5492-5493. Mohrig, J. R.; Keegstra, K.; Maverick, A.; Roberts, R.; Wells, S. J. Chem. Soc., Chem. Commun. 1974, 780-781.
- The site of protonation of α -diazo ketones is most probably the carbonyl oxygen. See the following. Avaro, M.; Levisalles, J.; Sommer, J. M. Chem. Commun. 1968, 410–412. Wentrup, C.; Dahn, H. Helv. Chim. Acta 1970, 53. 1637-1645.
- (3) McGarrity, J. F.; Smyth, T. J. Chem. Soc., Chem. Commun. 1977, 347-348. McGarrity, J. F.; Smyth, T. Abstracts of Papers of the Fourth IUPAC Symposium on Physical Organic Chemistry, York, England, 1978.
- The structure presented here is the most apparent that contains an sp2hybridized carbon in a CH₂ group, indicated by the spectroscopic data; no information is available on N–N bond lengths or N–N–H angles that might eliminate other structures.
- Pruckmayr, G.; Wu. T. K. Macromolecules 1975, 8, 954-956
- (6)Olah, G. A.; Sommer, J.; Namanworth, E. J. Am. Chem. Soc. 1967, 89, 3576-3586
- (7) Jacobsen, J. P.; Schaumburg, K.; Nielsen, J. T. J. Magn. Reson. 1975, 13, 372-378.
- Olah, G. A.; White, A. M. J. Am. Chem. Soc. 1969, 91, 5801-5810.
- Firl, J.; Runge, W.; Hartmann, W. Angew. Chem. **1974**, *86*, 274–275; Angew. Chem., Int. Ed. Engl. **1974**, *13*, 270–271.
- (10)We thank a referee for bringing this possibility to our attention.
- (11) Sample prepared and measured by Dr. D. P. Cox. (12) Ahmed, M. G.; Alder, R. W.; James, G. H.; Sincott, M. L.; Whiting, M. C. Chem. Commun. 1968, 1533-1534.
- Ohmen, R.; Schmitz, E. Chem. Ber. 1964, 97, 297–298.
 Duthalter, R. O.; Förster, H. G.; Roberts, J. D. J. Am. Chem. Soc. 1978, 100, 4974-4979.
- Runge, W.; Firl, J. Z. Naturforsch. B 1976, 31, 1515-1518. Albright, T. A.; Freeman, W. J. Org. Magn. Reson. 1977, 9, 75–79. (16) Niemeyer, H. M. Helv. Chim. Acta 1976, 59, 1133–1139. (17) Taft, R. W.; Wolf, J. F.; Beauchamp, J. L.; Scorrano, G.; Arnett, E. M. J. Am.
- (17)Chem. Soc. 1978, 100, 1240-1249.

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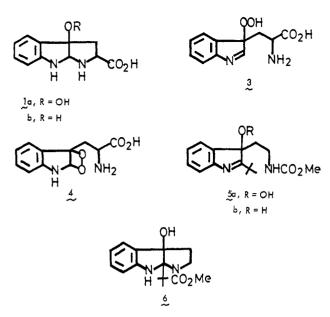
3a-Hydroperoxypyrroloindole from Tryptophan. Isolation and Transformation to Formylkynurenine

Sir:

Since the early 1950s, when the first report on Methylene Blue sensitized photooxygenation of tryptophan appeared,¹ this field of study has been developed as a tool for the elucidation of the active site in enzymes, and as a model reaction for the dioxygenase-catalyzed reaction of tryptophan to formylkynurenine.² There is, however, considerable variation in the earlier results and very little attention has been given to the reaction mechanism.

Recently we reported that Rose Bengal sensitized photooxygenation of tryptophan³ and tryptophan derivatives⁴ followed by Me₂S reduction gave 3a-hydroxypyrroloindoles,

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whereas formylkynurenine was isolated by heating the oxygenation mixture obtained from tryptophan. In this paper we report the first successful isolation, the nature, and the reactivity of the initially formed peroxidic intermediate in the dye-sensitized photooxygenation of tryptophan.

L-Tryptophan (1 g, 5 mM) was irradiated⁵ in 5% EtOH- H_2O containing Rose Bengal (15 mg, $\frac{1}{300}$ mol equiv) as sensitizer at 0-5 °C for 3.5 h, while oxygen was bubbled through the mixture, which was then acidified with AcOH and extracted with CH₂Cl₂ to remove the sensitizer. Lyophilization of the aqueous solution⁶ gave the tricyclic hydroperoxide 1a as a powder in 75-85% yield7 which was successfully purified on a Sephadex G-10 column to give a colorless powder. 1a: λ_{max} (H₂O) nm (ϵ), 235 (6300), 294.5 (2000); ν_{max} (KBr) 1610 cm^{-1} ; NMR δ (D₂O) 2.50-3.20 (m, 2 H, C₃ H), 4.01 (t, 0.6 H, cis C_2 H), 4.39 (t, 0.4 H, trans C_2 H), 5.64 (s, 0.4 H, trans NCHN), 5.75 (s, 0.6 H, cis NCHN), 6.70-7.15, 7.15-7.60 (m, 4 H, aromatic H); m/e 236 (M+). Chromatographic analysis on silica gel as well as the NMR spectrum revealed 1a to be a mixture of cis and trans isomers, reduction of which with Me₂S furnished the corresponding hydroxy derivatives 1b.8 The hydroperoxide 1a was partially converted to formylkynurenine (2, 21%) along with 1b (55%) when heated in water at 100 °C for 30 min. The aqueous solution of 1a was also found to convert to 2 (11%) and 1b (70%) after 1 week at room temperature. Decomposition of 1a was catalyzed by silica gel, metal ions such as Fe^{3+} and Fe^{2+} , and light (253.7 or >300 nm); however, cupric and cuprous ions did not show any significant effect.9 On the other hand, the exclusive formation of formylkynurenine (2, 64%) from 1a was observed when an aqueous solution of 1a was basified with Na₂CO₃ to about pH 9.10 In contrast, by acidification with HCl. 1a underwent a facile acid-catalyzed rearrangement followed by hydrolysis to give o-aminophenol in 40% yield.4c.c

The rearrangement of 1a to formylkynurenine could be explained by assuming an eight-membered unstable cyclic carbinol as an intermediate.^{4e} Yet there is another possibility involving 3 derived from 1a to give 2 via the dioxetane 4, although evidence for the existence of an equilibrium between 1a and 3 is lacking. In order to demonstrate tautomerism between 1a and 3, we have carried out the analogous photosensitized oxygenation of 2-*tert*-butyl- N^{b} -methoxycarbonyltryptamine which has the bulky substituent at the 2 position. As expected, the hydroperoxyindolenine **5a**,¹¹ corresponding to the putative primary intermediate **3** of the dye-sensitized photooxygenation of tryptophan and tryptamines, was isolated in 90% yield. Reduction of **5a** with Me₂S gave **5b**, mp 157-159 °C.¹² The NMR spectrum of **5b** showed that **5b** exists as a tautomeric mixture of **5b** and **6** (15:85 in CDCl₃, 4:1 in CD₃OD). Refluxing **5b** in CH₂Cl₂ followed by recrystallization from CH₂Cl₂-hexane gave **6**, mp 136-138 °C, which showed an NMR spectrum identical with that obtained from **5b** after equilibrium is reached.

These results, by reasons of analogy, may be regarded as strong evidence for the presence of an equilibrium between hydroperoxyindolenine 3 and the hydroperoxypyrroloindole 1a and support the mechanism previously proposed for the formation of formylkynurenine (2) from 1a via 3 and $4.^{3,13}$. These results provide a powerful predictive model for mechanism of biological oxidation of tryptophan.

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References and Notes

- (1) L. Weil, W. G. Gordon, and A. B. Buchert, Arch. Biochem. Biophys., 33, 90 (1951).
- (2) A Fontana and C. Toniolo, Fortschr. Chem. Org. Naturst. 33, 309–449 (1976); M. Nakagawa and T. Hino, Yuki Gosei Kagaku Kyokai Shi, 35, 42 (1977).
- M. Nakagawa, H. Watanabe, S. Kodato, H. Okajima, T. Hino, J. L. Flippen, and B. Witkop, *Proc. Natl. Acad. Sci. U.S.A.*, 74, 4730 (1977).
 (4) (a) M. Nakagawa, T. Kaneko, K. Yoshikawa, and T. Hino, *J. Am. Chem. Soc.*.
- (4) (a) M. Nakagawa, T. Kaneko, K. Yoshikawa, and T. Hino, J. Am. Chem. Soc., 96, 624 (1974); (b) M. Nakagawa, K. Yoshikawa, and T. Hino, *ibid.*, 97, 6496 (1975); (c) M. Nakagawa, H. Okajima, and T. Hino, *ibid.*, 98, 635 (1976); (d) M. Nakagawa, H. Okajima, and T. Hino, *ibid.*, 99, 4424 (1977); (e) I. Saito, T. Matsuura, M. Nakagwa, and T. Hino, Acc. Chem. Res., 10, 346 (1977); (f) M. Nakagawa, J. Chiba, and T. Hino, *Heterocycles*, 9, 385 (1978).
- (5) A 500-W halogen lamp with CuCl2-CaCl2 filter was used.
- (6) Neither formylkynurenine nor N^b-formylkynurenine has been detected (TLC, UV spectrum).
- (7) The yield was determined by iodometry. The extent of decomposition of 1a was shown to be <5% when kept at about -70 °C for 2 months, while 1a decomposes at room temperature within 24 h.
- (8) The L-series 1b showed UV and NMR spectra identical with those of the DL series 1b. The stereochemistry of the two isomers of 1b has been established.³ Cf. W. E. Savige, Aust. J. Chem., 28, 2275 (1975).
- (9) Decomposition was monitored by iodometry as well as TLC. For an example, when FeCl₃ (0.1 mol equiv) was added, 1a was decomposed after 24 h. Acid hydrolysis of the reaction mixture with F_3CCO_2H followed by Dowex 50 \times 8 column separation afforded kynurenine (2%) and 1b (59%).
- (10) The mechanism of the base-catalyzed conversion is not clear. However, the reverse reaction of 1a to 3 might be catalyzed by the base. The dioxetane formation or addition of water to the C = N double bond of the indolenine 3 may be favored in alkaline conditions.
- (11) 5a: chromatographically homogeneous oil; λ_{max} (EtOH), nm, 222, 264, 292 (sh); ν_{max} (KBr), cm⁻¹, 1700, 1560, 1535; δ (CDCl₃) 1.44 (s, 9 H, Me₃C), 2.00–2.44 (m, 2 H, CH₂), 2.44–3.30 (m, 2 H, CH₂N), 3.56 (s, 3 H, CO₂CH₃); m/ e 306 (M⁺). 5a was converted to the formylkynurenine analogue, mp 120.5–122.5 °C, when refluxed in MeOH.
- (12) Satisfactory elemental analysis and spectral data were obtained for all new compounds characterized by melting points.
 (13) A weak chemiluminescence was observed in a dark room when 1a was
- (13) A weak chemiluminescence was observed in a dark room when 1a was heated in Me₂SO at 170 °C.

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